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A DNA encoding melon phytoene synthase.

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DNA, CONSTRUCTS, CELLS AND PLANTS DERIVED THEREFROM

This application relates to novel DNA constructs, plant cells containing the constructs and plants derived therefrom. In particular it relates to the modification of plant gene expression causing modification of carotenoid metabolism in plants.

The modification of plant gene expression has been achieved by several methods. The molecular biologist can choose from a range of known methods to decrease or increase gene expression or to alter the spatial or temporal expression of a particular gene. For example, the expression of either specific antisense RNA or partial (truncated) sense RNA has been utilised to reduce the expression of various target genes in plants (as reviewed by Bird and Ray, 1991, *Biotechnology and Genetic Engineering Reviews* 9:207-227). These techniques involve the incorporation into the genome of the plant of a synthetic gene designed to express either antisense or sense RNA. They have been successfully used to down-regulate the expression of a range of individual genes involved in the development and ripening of tomato fruit (Gray et al, 1992, *Plant Molecular Biology*, 19:69-87). Methods to increase the expression of a target gene have also been developed. For example, additional genes designed to express RNA containing the complete coding region of the target gene may be incorporated into the genome of the plant to "over-express" the gene product. Various other methods to modify gene expression are known; for example, the use of alternative regulatory sequences.

Carotenoids include a range of biochemicals such as carotenes, lutein, xanthophylls, and pigments such as lycopene. The biosynthesis of carotenoids in higher plants and micro-organisms has been the subject of numerous investigations. The pathways of carotenoid biosynthesis across a range of organisms share many common features (Bramley and Mackenzie, 1988). The biochemical pathway leading to carotenoid biosynthesis is shown in Figure 1.

The divergence to the dedicated pathway for carotenoid synthesis is after the formation of geranylgeranyl diphosphate (GGDP - previously known as geranylgeranyl pyrophosphate, GGPP). The first unique step in the formation of carotenoids is the head-to-head condensation of 2 molecules of

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GGDP to form the first C₄₀ carotenoid, phytoene. The synthesis of phytoene from GGDP involves two reaction steps with prephytoene diphosphate (PPDP - previously known as prephytoene pyrophosphate, PPPP) as the intermediate, but both reactions are catalysed by a single enzyme in plants and micro-organisms. This enzyme, which catalyses the conversion of GGDP to phytoene, is known as phytoene synthase (Bramley et al, 1992, Plant Journal, 2:343-349).

Phytoene synthase activity has been detected in a wide range of plant species including tomato, pepper, maize, carrot, spinach and tobacco. There are many similarities between the enzymes from different species. The enzymes are located in the plastid stroma and synthesise the same phytoene stereo-isomer (15-cis phytoene). The enzymes all require Mn⁺⁺ as a co-factor although the requirement for ATP is unclear.

DNA sequences encoding phytoene synthase enzymes are known. For example, the TOM5 gene encodes an enzyme involved in the ripening of tomatoes (Ray et al, 1987, Nucleic Acids Research, 15:10587) which has been identified as the enzyme phytoene synthase (international patent application published as W091/09128; international patent application published as W092/16635). Phytoene synthase enzymes show sequence similarity. The molecular weight of the purified pepper (Capsicum annuum) phytoene synthase (48kD) (described by Dogbo et al 1988) is very similar to the predicted molecular weight of the mature phytoene synthase protein encoded by TOM5 (47.6kD). Alignment of the predicted phytoene synthase protein sequences from Arabidopsis and pepper cDNA clones with the predicted protein sequence of the tomato phytoene synthase encoded by TOM5 shows 75% and 90.6% identity respectively. The protein encoded by the tomato TOM5 cDNA also shows significant homology (32% identity; 56.5% similarity) to the CrtB protein from Rhodobacter capsulatus, a gram-negative purple bacterium (Armstrong et al, 1990, JBC, 265:8329-8338). The CrtB protein catalyses the conversion of GGDP to phytoene (Bartley GE et al, 1992, J Biological Chem, 267:5036-5039).

There is homology between plant phytoene synthase DNA sequences. Tomato contains at least one other locus with homology to TOM5. TOM5 cDNA will cross-hybridise with two clones: the genomic clone known as Clone F (Ray et al, 1992, PMB, 19:401-404) and the cDNA clone known as Psy2 (Bartley and Scolnik, 1993, J Biol Chem, 268:25718-25721). Psy2 has 80%

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overall nucleotide homology to TOM5 cDNA. The TOM5 cDNA sequence has also been used as a hybridisation probe to identify phytoene synthase clones from other species, including Capsicum annuum which has 83% overall homology (Romer et al, 1993, Biochem Biophys Res Commun, 196:1414-1421) and Arabidopsis thaliana which has 65% overall homology (Scolnik and Bartley, 1994, Plant Physiology, 104:1471-1472).

International application W091/09128 (EP-A-505405) describes "antisense" and "sense" DNA constructs encoding phytoene synthase and their use to modify the synthesis of carotenoid compounds in plants. These DNA constructs comprise a DNA sequence homologous to some or all of the TOM5 gene, preceded by a transcriptional initiation region operative in plants so that the construct can generate mRNA in plant cells. The DNA construct may encode mRNA which can be translated to give the enzyme produced by the TOM5 gene. International application W091/09128 describes a method for modifying (inhibiting or promoting) the production of carotenoids in plants by transforming such plants with the aforementioned DNA constructs, and also describes transformed plants (and their descendants) having a modified carotenoid content. Generally, production of the TOM5 enzyme (phytoene synthase) within the plant is enhanced by constructs which contain DNA homologous to the substantially complete gene. For example, promotion of lycopene production may be brought about by inserting one or more functional copies of the TOM5 cDNA, or of the full-length TOM5 gene, under control of a functional plant promoter into a tomato plant to give fruit of a deeper red colour. Fray and Grierson (1993, Plant Mol Biol, 22:589-602) discuss the over-expression of plant phytoene synthase genes in tomato and the effect on colour.

In summary, international application W091/09128 describes the control of TOM5 gene expression in plants (using recombinant DNA technology) to modify carotenoid metabolism (by increasing or decreasing total carotenoid content of plant parts or increasing/decreasing levels of individual carotenoids). In particular, it describes the modification of colour production and certain other related functions (β -carotene and Vitamin A production; protection against high light intensity damage; control of sporopollenin synthesis and pollen formation/maturation).

In work leading to the present invention we have identified an alternative DNA sequence encoding a phytoene synthase which is useful for

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modifying carotenoid metabolism in plants.

According to the present invention, there is provided a DNA sequence encoding melon phytoene synthase. The DNA sequence may be derived from cDNA, from genomic DNA or may be synthesised ab initio.

cDNA clones encoding melon phytoene synthase may be obtained from melon cDNA libraries. Sequences coding for the whole, or substantially the whole, of the mRNA produced by the corresponding gene may thus be obtained. The cDNA so obtained may be sequenced according to known methods.

A cDNA clone encoding phytoene synthase has been isolated from melon. The clone (hereinafter referred to as MEL5) is almost full length and its nucleic acid sequence (with derived amino acid sequence) is shown in Sequence ID-1.

An alternative source of the DNA sequence is a suitable gene encoding melon phytoene synthase. This gene may differ from the corresponding cDNA in that introns may be present. The introns are not transcribed into mRNA (or, if so transcribed, are subsequently cut out). Oligonucleotide probes or the cDNA clone may be used to isolate the actual phytoene synthase gene(s) by screening melon genomic DNA libraries. Such genomic clones may include control sequences operating in the plant genome. Thus it is also possible to isolate promoter sequences which may be used to drive expression of the enzymes or any other protein. These promoters may be particularly responsive to certain developmental events and environmental conditions. The melon phytoene synthase gene promoters may be used to drive expression of any target gene.

A further way of obtaining a melon phytoene synthase DNA sequence is to synthesise it ab initio from the appropriate bases, for example using the appropriate cDNA sequence as a guide.

Some or all of the melon phytoene synthase sequence may be incorporated into DNA constructs suitable for plant transformation. These DNA constructs may then be used to modify gene expression in plants. "Antisense" or "partial sense" or other techniques may be used to reduce the expression of the phytoene synthase in plant tissue. The levels of phytoene synthase may also be increased; for example, by incorporation of additional enzyme genes. The additional genes may be designed to give either the same or different spatial and temporal patterns of expression in the plant.

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According to a further aspect of the invention there is provided a DNA construct comprising some or all of a DNA sequence encoding melon phytoene synthase under the control of a transcriptional initiation region operative in plants, so that the construct can generate RNA in plant cells.

The carotenoid content and related characteristics of plant parts (particularly fruit) may be modified by transformation with a DNA construct according to the invention. The invention also provides plant cells containing such constructs; plants derived therefrom showing modified carotenoid content; and seeds of such plants.

A DNA construct according to the invention may be an "antisense" construct generating "antisense" RNA or a "sense" construct (encoding at least part of the functional enzyme) generating "sense" RNA. "Antisense RNA" is an RNA sequence which is complementary to a sequence of bases in the corresponding mRNA: complementary in the sense that each base (or the majority of bases) in the antisense sequence (read in the 3' to 5' sense) is capable of pairing with the corresponding base (G with C, A with U) in the mRNA sequence read in the 5' to 3' sense. Such antisense RNA may be produced in the cell by transformation with an appropriate DNA construct arranged to generate a transcript with at least part of its sequence complementary to at least part of the coding strand of the relevant gene (or of a DNA sequence showing substantial homology therewith). "Sense RNA" is an RNA sequence which is substantially homologous to at least part of the corresponding mRNA sequence. Such sense RNA may be produced in the cell by transformation with an appropriate DNA construct arranged in the normal orientation so as to generate a transcript with a sequence identical to at least part of the coding strand of the relevant gene (or of a DNA sequence showing substantial homology therewith). Suitable sense constructs may be used to inhibit gene expression (as described in International Patent Publication W091/08299) or a sense construct encoding and expressing the functional enzyme may be transformed into the plant to over-express the enzyme.

DNA constructs according to the invention may comprise a base sequence at least 10 bases (preferably at least 35 bases) in length for transcription into RNA. There is no theoretical upper limit to the base sequence - it may be as long as the relevant mRNA produced by the cell - but for convenience it will generally be found suitable to use sequences

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between 100 and 1000 bases in length for downregulation purposes but somewhere in excess of about 1500 bases is preferred for overexpression. The preparation of such constructs is described in more detail below.

As a source of the DNA base sequence for transcription, a suitable cDNA or genomic DNA or synthetic polynucleotide may be used. The isolation of suitable melon phytoene synthase-encoding sequences is described above. Sequences coding for the whole, or substantially the whole, of the enzyme may thus be obtained. Suitable lengths of this DNA sequences may be cut out for use by means of restriction enzymes. When using genomic DNA as the source of a partial base sequence for transcription it is possible to use either intron or exon regions or a combination of both.

To obtain constructs suitable for expression of melon phytoene synthase in plant cells, the cDNA sequence as found in the enzyme cDNA or the gene sequence as found in the chromosome of the plant may be used. Recombinant DNA constructs may be made using standard techniques. For example, the DNA sequence for transcription may be obtained by treating a vector containing said sequence with restriction enzymes to cut out the appropriate segment. The DNA sequence for transcription may also be generated by annealing and ligating synthetic oligonucleotides or by using synthetic oligonucleotides in a polymerase chain reaction (PCR) to give suitable restriction sites at each end. The DNA sequence is then cloned into a vector containing upstream promoter and downstream terminator sequences. If antisense DNA is required, the cloning is carried out so that the cut DNA sequence is inverted with respect to its orientation in the strand from which it was cut.

In a construct expressing antisense RNA, the strand that was formerly the template strand becomes the coding strand, and vice versa. The construct will thus encode RNA in a base sequence which is complementary to part or all of the sequence of the enzyme mRNA. Thus the two RNA strands are complementary not only in their base sequence but also in their orientations (5' to 3').

In a construct expressing sense RNA, the template and coding strands retain the assignments and orientations of the original plant gene. Constructs expressing sense RNA encode RNA with a base sequence which is homologous to part or all of the sequence of the mRNA. In constructs which express the functional enzyme, the whole of the coding region of the gene

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is linked to transcriptional control sequences capable of expression in plants.

For example, constructs according to the present invention may be made as follows. A suitable vector containing the desired base sequence for transcription (such as the MEL5 cDNA clone) is treated with restriction enzymes to cut the sequence out. The DNA strand so obtained is cloned (if desired, in reverse orientation) into a second vector containing the desired promoter sequence and the desired terminator sequence. Suitable promoters include the 35S cauliflower mosaic virus promoter and the tomato polygalacturonase gene promoter sequence (Bird et al, 1988, Plant Molecular Biology, 11:651-662) or other developmentally regulated fruit promoters. Suitable terminator sequences include that of the Agrobacterium tumefaciens nopaline synthase gene (the nos 3' end).

In a DNA construct according to the invention, the transcriptional initiation region may be derived from any plant-operative promoter. The transcriptional initiation region may be positioned for transcription of a DNA sequence encoding RNA which is complementary to a substantial run of bases in a mRNA encoding phytoene synthase (making the DNA construct a full or partial antisense construct).

The transcriptional initiation region (or promoter) operative in plants may be a constitutive promoter (such as the 35S cauliflower mosaic virus promoter) or an inducible or developmentally regulated promoter (such as fruit-specific promoters), as circumstances require. For example, it may be desirable to modify enzyme activity only during fruit development and/or ripening. Use of a constitutive promoter will tend to affect enzyme levels and functions in all parts of the plant, while use of a tissue specific promoter allows more selective control of gene expression and affected functions (eg fruit colouration). Thus in applying the invention it may be found convenient to use a promoter that will give expression during fruit development and/or ripening. Thus the antisense or sense RNA is only produced in the organ in which its action is required. Fruit development and/or ripening-specific promoters that could be used include the ripening-enhanced polygalacturonase promoter (International Patent Publication Number W092/08798), the E8 promoter (Diekman & Fischer, 1988, EMBO, 7:3315-3320) and the fruit specific 2A11 promoter (Pear et al, 1989, Plant Molecular Biology, 13:639-651).

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The DNA constructs of the invention may be inserted into plants to regulate the production of phytoene synthase resulting in modification of carotenoid metabolism. Depending on the nature of the construct, the production of the enzyme may be increased, or reduced, either throughout or at particular stages in the life of the plant. Generally, as would be expected, production of the enzyme is enhanced only by constructs which express RNA homologous to the substantially complete endogenous enzyme mRNAs. Full-length sense constructs may also inhibit enzyme expression. Constructs containing an incomplete DNA sequence shorter than that corresponding to the complete gene generally inhibit the expression of the gene and production of the enzymes, whether they are arranged to express sense or antisense RNA.

A DNA construct of the invention is transformed into a target plant cell. The target plant cell may be part of a whole plant or may be an isolated cell or part of a tissue which may be regenerated into a whole plant. The target plant cell may be selected from any monocotyledonous or dicotyledonous plant species.

Constructs according to the invention may be used to transform any plant using any suitable transformation technique to make plants according to the invention. Both monocotyledonous and dicotyledonous plant cells may be transformed in various ways known to the art. In many cases such plant cells (particularly when they are cells of dicotyledonous plants) may be cultured to regenerate whole plants which subsequently reproduce to give successive generations of genetically modified plants. Any suitable method of plant transformation may be used. For example, dicotyledonous plants such as tomato and melon may be transformed by Agrobacterium Ti plasmid technology, such as described by Bevan (1984, Nucleic Acid Research, 12:8711-8721) or Fillatti et al (Biotechnology, July 1987, 5:726-730). Such transformed plants may be reproduced sexually, or by cell or tissue culture.

Examples of genetically modified plants according to the present invention include all fruit-bearing plants (such as tomatoes, mangoes, peaches, apples, pears, strawberries, bananas, melons, peppers, chillies, paprika). For example, the fruit of such plants may be made more attractive (or at least interesting) by inducing or intensifying a red colour therein. Other plants that may be modified by the process of the

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invention include tubers such as radishes, turnips and potatoes, as well as cereals such as maize (corn), wheat, barley and rice. Flowers and ornamental grasses may also be modified.

Use of the novel melon phytoene synthase constructs allows modification of enzyme activity and thus provides a method for modification of carotenoid biosynthesis. The overall level of phytoene synthase activity and the relative activities of the individual enzymes of the carotenoid pathway affect the development and final form of carotenoid content in the plant and thus determine certain characteristics of the plant parts. Modification of phytoene synthase activity can therefore be used to modify various aspects of plant (including fruit) quality. The activity levels of the phytoene synthase enzyme may be either increased or reduced during development depending on the characteristics desired for the modified plant. Enhancing expression of phytoene synthase will tend to increase production of certain carotenoids, and inhibiting expression will tend to decrease production. Phytoene synthase may also be expressed in cells, tissues and organisms that do not normally produce the enzyme.

For example, the down-regulation or over-expression of phytoene synthase may be used to modify (alter or enhance) fruit colour by altering pigment levels. This is particularly useful for modifying fruit and vegetable colour but may equally be applied to leaves and other organs.

Carotenoid content (and hence plant characteristics) may be modified to a greater or lesser extent by controlling the degree of the sense or antisense mRNA production in the plant cells. This may be done by suitable choice of promoter sequences, or by selecting the number of copies or the site of integration of the DNA sequences that are introduced into the plant genome. For example, the DNA construct may include more than one DNA sequence encoding phytoene synthase or more than one recombinant construct may be transformed into each plant cell.

It is also possible to modify the activity of phytoene synthase while also modifying the activity of one or more other enzymes. For example, the other enzymes may be involved in cell metabolism or in fruit development and ripening. Cell wall metabolising enzymes that may be modified in combination with phytoene synthase include but are not limited to: pectin esterase, polygalacturonase, β -galactanase, β -glucanase. Other enzymes involved in fruit development and ripening that may be modified in

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combination with phytoene synthase include but are not limited to: ethylene biosynthetic enzymes, other carotenoid biosynthetic enzymes, carbohydrate metabolism enzymes including invertase.

Several methods are available for modification of the activity of phytoene synthase in combination with other enzymes. For example, a first plant may be individually transformed with a phytoene synthase construct and then crossed with a second plant which has been individually transformed with a construct encoding another enzyme. As a further example, plants may be either consecutively or co-transformed with phytoene synthase constructs and with appropriate constructs for modification of the activity of the other enzyme(s). An alternative example is plant transformation with a phytoene synthase construct which itself contains an additional gene for modification of the activity of the other enzyme(s). The phytoene synthase constructs may contain sequences of DNA for regulation of the expression of the other enzyme(s) located adjacent to the phytoene synthase sequences. These additional sequences may be in either sense or antisense orientation as described in International patent application publication number W093/23551 (single construct having distinct DNA regions homologous to different target genes). By using such methods, the benefits of modifying the activity of phytoene synthase may be combined with the benefits of modifying the activity of other enzymes.

According to a third aspect of the invention, there is provided a method for modifying the production of carotenoids in plants by transforming such plants with DNA adapted to modify carotenoid biosynthesis and growing such transformed plants or their descendants followed by selection of plants having plant parts (for example leaves, petals or fruit) of modified carotenoid content. Suitable DNA comprises constructs according to the invention which may be adapted to enhance the production of one or more carotenoids or to inhibit such production by the plant.

In this way, plants can be generated which have modified colour due to promotion or inhibition of the pathways of carotenoid biosynthesis. In particular, phytoene synthase constructs may be used to promote or inhibit the production of the red colour associated with lycopene. Similar colour modifications may be possible using traditional plant breeding techniques, but the present invention provides a means of transferring the trait into elite lines without a prolonged breeding programme which might alter other

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traits at the same time. Promotion of lycopene production (eg by sense over-expression constructs) may produce fruit of a deeper red colour, which may appear more appetising to the consumer. The invention may also be used to introduce a red colour into parts of plants other than the fruit. For example, promotion of lycopene may be brought about by inserting one or more functional copies of the gene cDNA, or of the full-length gene, under control of a promoter functional in plants. If phytoene synthase is naturally expressed in the plant, the promoter may be selected to give a higher degree of expression than is given by the natural promoter.

As well as colour production, other important functions may be modified by the process of the invention. Thus β -carotene (a precursor of Vitamin A) and other carotenoids are important to human health, and have been claimed to have a protective effect against certain diseases. Food plants may be modified by transformation with the constructs of the invention so that they have a higher content of such compounds: or other plants may be so modified, so that they can act as a source from which such compounds can be extracted. Carotenoids are also believed to have a role in protecting plants against high light intensity damage, so plants with a higher content of such compounds may be of value in combating the effects of any global climate change.

As already discussed, plants produced by the process of the invention may also contain other recombinant constructs, for example constructs having other effects on fruit ripening. For example fruit of enhanced colour according to the invention may also contain constructs inhibiting the production of enzymes such as polygalacturonase and pectinesterase, or interfering with ethylene production. Fruit containing both types of recombinant construct may be made either by successive transformations, or by crossing two varieties that each contain one of the constructs, and selecting among the progeny for those that contain both.

The invention will now be described by way of example only, with reference to the drawings in which:

Figure 1 is a diagram of the carotenoid biosynthesis pathway;

Figure 2 illustrates a sense construct; and,

Figure 3 illustrates an antisense construct.

The following gene sequences are provided herewith:

Sequence ID-1 is the nucleotide sequence of MEL5;

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Sequence ID-2 is the sequence of TOM 5, the equivalent sequence from tomato.

EXAMPLE 1

cDNA clone encoding melon phytoene synthase

The MEL5 cDNA clone was isolated from a ripening related cDNA library derived from climacteric melon fruit, using the tomato phytoene synthase cDNA (TOM5, see Ray et al, 1992 or EP-A-505405) as a heterologous probe. Genomic Southern analysis indicated that the MEL5 clone is encoded by a single copy or small number of genes (genomic melon DNA was digested with BamHI, EcoRV, BamHI/EcoRV and hybridised to radiolabelled MEL5 insert, followed by washing up to 0.1xSSC, 0.1% SDS for 30 minutes at 42°C or 50°C).

Northern analysis of RNA extracted from a ripening series of fruit and from roots, leaves and flower petals was used to examine the pattern of expression of this mRNA. Total RNA was extracted from different ripening stages of fruit (green fruit not producing ethylene; green/orange fruit producing 5-7 nl/g/h ethylene; orange fruit producing 3-6 nl/g/h and very orange fruit producing approximately 1 nl/g/h ethylene). Total RNA and poly(A)⁺ RNA from fruit, leaves, petals and roots was hybridised to radiolabelled MEL5 insert. MEL5 hybridised to a transcript of 1.7 kB which was present at all stages of fruit ripening, in petals and in trace amounts in leaves and roots. The cDNA was 1.36 kB in size, and sequence analysis of the longest open reading frame demonstrated 68% identity at the nucleotide level and 83.1% similarity at the amino acid level with the tomato fruit phytoene synthase. The MEL5 cDNA clone was not full length but the 5' end was isolated using a PCR approach, and subsequently fully sequenced (Sequence ID-1).

EXAMPLE 2

Construction of antisense RNA vectors with the CaMV 35S promoter

A vector is constructed using the sequences corresponding to a fragment of the insert of a melon phytoene synthase cDNA. This fragment is synthesised by polymerase chain reaction using synthetic primers. The ends of the fragment are made flush with T4 polymerase and it is cloned into the vector pJR1 which has previously been cut with SmaI. pJR1 (Smith et al, 1988, Nature, 334:724-726) is a Bin19 (Bevan, 1984, Nucleic Acids Research, 12:8711-8721) based vector, which permits the expression of the antisense

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RNA under the control of the CaMV 35S promoter. This vector includes a nopaline synthase (nos) 3' end termination sequence.

Alternatively a vector is constructed using a restriction fragment obtained from a melon phytoene synthase cDNA and cloned into the vectors GA643 (An *et al*, 1988, Plant Molecular Biology Manual A3: 1-19) or pDH51 (Pietrzak *et al*, 1986, Nucleic Acids Research, 14:5875-5869) which has previously been cut with a compatible restriction enzyme(s). A restriction fragment from the phytoene synthase/pDH51 clone containing the promoter, the phytoene synthase fragment and other pDH51 sequence is cloned into SLJ44026B or SLJ44024B (Jones *et al*, 1990, Transgenic Research, 1) or Bin19 (Bevan, 1984, Nucleic Acids Research, 12:8711-8721) which permits the expression of the antisense RNA under control of the CaMV 35S promoter. This procedure is illustrated in Figures 2 and 3.

After synthesis of the vector, the structure and orientation of the sequences are confirmed by DNA sequence analysis.

EXAMPLE 3

Construction of antisense RNA vectors with a fruit enhanced promoter.

Referring to Figures 2 and 3 the fragment of the melon phytoene synthase cDNA that was described in Example 2 is also cloned into the vector pJR3. pJR3 is a Bin19 based vector, which permits the expression of the antisense RNA under the control of the tomato polygalacturonase (PG) promoter. This vector includes approximately 5 kb of promoter sequence and 1.8 kb of 3' sequence from the PG promoter separated by a multiple cloning site.

After synthesis, vectors with the correct orientation of the phytoene synthase sequences are identified by DNA sequence analysis.

Alternative fruit enhanced promoters (such as E8 or 2A11) are substituted for the polygalacturonase promoter in pJR3 to give alternative patterns of expression.

EXAMPLE 4

Construction of truncated sense RNA vectors with the CaMV 35S promoter

The fragment of the phytoene synthase cDNA that was described in Example 2 is also cloned into the vectors described in Example 2 in the sense orientation.

After synthesis, the vectors with the sense orientation of the phytoene synthase sequence are identified by DNA sequence analysis.

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EXAMPLE 5

Construction of truncated sense RNA vectors with fruit-enhanced promoter.

The fragment of the phytoene synthase cDNA that was described in Example 3 is also cloned into the vectors described in Example 3 in the sense orientation.

After synthesis, the vectors with the sense orientation of the phytoene synthase sequence are identified by DNA sequence analysis.

EXAMPLE 6

Construction of an over-expression vector using the CaMV35S promoter

The complete sequence of a melon phytoene synthase cDNA containing a full open-reading frame is inserted into the vectors described in Example 2.

EXAMPLE 7

Construction of an over-expression vector using a fruit-enhanced promoter

The complete sequence of a melon phytoene synthase cDNA containing a full open-reading frame is inserted into the vectors described in Example 3 (pJR3 or alternatives with different promoters).

EXAMPLE 8

Generation of transformed plants

Vectors are transferred to Agrobacterium tumefaciens LBA4404 (a micro-organism widely available to plant biotechnologists) and are used to transform tomato plants. Transformation of tomato stem segments follow standard protocols (e.g. Bird et al, 1988, Plant Molecular Biology, 11:651-662). Transformed plants are identified by their ability to grow on media containing the antibiotic kanamycin. Plants are regenerated and grown to maturity. Ripening fruit are analyzed for modifications to their carotenoid content.

- 15 -

51 AAGAGAGGGTGGAAATTTGTAAGTCAAGAAACAGGTTACTCCTGTTTGAGT 200
1 ...TCCCTCGTGGGATACAAAAGAGAAGCTAAGTGGGAGT...TTCATCCGT 46
101 GAGGAAAAGTTGGTTTGCCTGTCTGTGGTCTTTTATAATCTTTTCTAC 250
47 GTCGAAGCATTTGCTTGGAAATTTCTTGGGCGGTTTAATTGGATTT..... 90
151 AGAAGAGAAAAGTGGGTAATTTGTTTGAGAGTGGAAATATTCTCTAGTGG 200
91 ...AAGTTATCTTGGCCTTTCCATTTTCATTTTGCAACTGAGTTTCTCGATT 138
201 GAATCTACTAGGAGTAATTTATTTTCTATAAACTAAGTAAAGTTTGAAG 250
139 CCTCCTGCGATTCTGTTCAATAAAGCCTTGGAAATCCTTCTAGTGGGAATA 188
251 GTGACAAAAGAAAGACAAAATCTTGGAAATGTTTTAGACAACCAAGGT 300
189 AAATCGATTGATATGCTTTAGCTTCATCGTTGGTTGTTTCTTCCAACGT 238
301 TTTCTTGCTCAGAATGTCTGTTCCTTGTATGGGTTGTTTCTCCTTGTG 350
239 TGAACCTTCCCA.....TCCAGCTTTGGGTTTCTTGATTGATTGAGAG 283
351 ACGTCTCAAATGGGACAAGTTTCATGGAATCAGTCCGGGAGGGAAACCGT 400
284 TGGACCCCAAATTCGGGATTCTTTAGATTCTCTTCGAGAAATCGAGTGC 333
401 TTTTGTGATTTCATCGAGGCATAGGAATTTGGTGTCCAATGAGAGAATCAA 450
334 CGAATCTGATTACAAAGAAACAGAAATGGGGGAATCATTCTCACTCTACA 383
451 TAGAGGTGGT..GGAAAGCAAACATAAATGGACGGAAATTTCTGTACG 498
384 GAACGAAATACCCAATTCTCCATGAAAGTGGATATGGGTCTGTTATTGT 433
499 GTCTGCTATTTTGGCTACTCCATCTGGAGAACGGACGATGACATCGGAAC 548
434 AGCAAGTATGGTGGCGAATCCCGCCGGAGAAATAGCCGTCCTCAGCTGAGC 483
549 AGATGGTCTATGATGTGGTTTGGAGGCAGGCAGCCTTGGTGAAGAGGCAA 598
484 AGAAGGTGTATAACGTAGTTATGAAACAGGCGGCTCTGGTGAACGACAA 533
599 CTGAGATCTACCAATGAGTTAGAAGTGAAGCCGGATATACCTATTCCGGG 648
534 CTTAGAACCSCCGGAGAATTGGATGTGAAGCCGGATATCGTTCTTCCGGG 583
649 GAATTTGGGCTTGTGAGTGAAGCATATGATAGGTGTGGTGAAGTATGTG 698
584 GACTTTGAGCTTGTGAATGAGGCTTATGATCGTGTGGTGAAGTTTGTG 633
599 CAGAGTATGCAAGACGTTTAACTTAGGAACTATGCTAATGACTCCCGAG 748
634 CAGAGTATGCCAAGACATTTTATCTGGGAATATGTTGATGACACCGGAG 683
749 AGAAGAAGGGCTATCTGGGCAATATATGTATGGTGCAGAAGAACAGATGA 798
684 AGGCAAAAGGCTATTGGGCAATTATGTATGGTGTAGGCGGACAGATGA 733
799 ACTTGTGATGGCCCAAACGCATCATATATTACCCGGCAGCCTTAGATA 848
734 ACTTGTGATGGGCCAAATGCTTCACATAACACCTACTGCATTGGACA 783

549 GGAGGGAAAATAGGCTAGAAATGTTTTCTATGGGCGGCCATTTGACATG 898
784 GATGGGAGGCAAGGCTGGAAGAGCTTTTCCAGGGAGGCCATTCCGATATG 833
899 CTCGATGGTGCTTTTGTCCGATACAGTTTCTAACTTTCCAGTTGATATTCA 948
934 CTCGATGCAGCTTTTGGCGGATCTGTTACTAAGTTCCCTGTGATATTCA 883
949 GCCATTCCAGAGATATGATTGAAGGAATGCGTATGGACTTGAGAAAATCGA 998
984 GCCGTTCAAAGATATGATTGAAGGGATGCGGATGGATCTAAGGAAGTCCA 933
999 GATACAAAACCTCGACGAACTATACCTTTATTTGTTATTATGTTGCTGGT 1048
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1049 ACGGTTGGGTTGATGAGTGTTCCTAATTATGGGTATCGCCCCCTGAATCAA 1098
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1039 GGCAACAACAGAGAGCGTATATAATGCTGCTTTGGCTCTGGGGATCGCAA 1148
1034 AGCAAGCACAGAGAGTGTGTACAATGCTGCCTTAGCATTAGGCATTGCCA 1083
1149 ATCAA...TTAACTAACATACTCAGAGATGTTGGAGAAGATGCCAGAAGA 1195
1084 ATCAAGCTCCACCAACATTCTCAGAGATGTTGGAGAAGATGCTAGAAGA 1133
1196 GGAAGAGTCTACTTGCTCAAGATGAATTAGCACAGGCAGGTCTATCCGA 1245
1134 GGAAGAATATATCTACCACAAGATGAGCTAGCACAGGCAGGTCTTTCAGA 1183
1246 TGAAGATATATTTGCTGGAAGGGTGACCGATAAATGGAGAATCTTTATGA 1295
1184 TGAGGACATATTTGCTGGAAGAGTAACTGATAAATGGAGAACTTCATGA 1233
1296 AGAAACAAATACATAGGGCAAGAAAGTTCCTTGATGAGGCAGAGAAAGGC 1345
1234 AGAATCAGATTAAGAGGGCTAGAATGTTCTTGATGAGGCTGAGAAAGGA 1283
1346 GTGACAGAATTGAGCTCAGCTAGTAGATTCCCTGTATGGGCATCTTTGGT 1395
1284 GTTTTGGAGCTTAATAAAGCTAGCAGATGGCCGGTGTGGGCTTCTTTGCT 1333
1396 CTGTACCGCAAAATACTAGATGAGATTGAAGCCAATGACTACAACAAC 1445
1334 ATTGTATAGGCAAAATATTGGATGAGATCGAAGCAAACGACTACGACAAC 1383
1446 TCACAAAGAGAGCATATGTGAGCAAATCAAAGAAGTTGATTGCATTACCT 1495
1384 TCACAAAAGGGCTTATGTGAGCAAAGCCAGAAAAATATTGGCTTTGCCA 1433
1496 ATTGCATATGCAAAATCTCTTGGCTCTTACAAAA...CTGCCCTCTCT 1542
1434 ATGGCTTATGGAAGGGCCCTCTCTTGGCTCTTCAATCTATACCACTAT 1483
1543 TCAAAGATAAAGCATGAATGAAGATATATATATATATATATAGCAA 1591
1484 ACACTCAACAACCAATCAAAATCAACCAAAAAAAGAAAAAAAT 1533
1592 TATACATTAGAAGAAAAAAGGAAGAAGAAATGTTGTTGTTATGATATAA 1641
1534 TGTATATTAGCATATTCTTATCTAATACAATGTCTCTCAATT..... 1576

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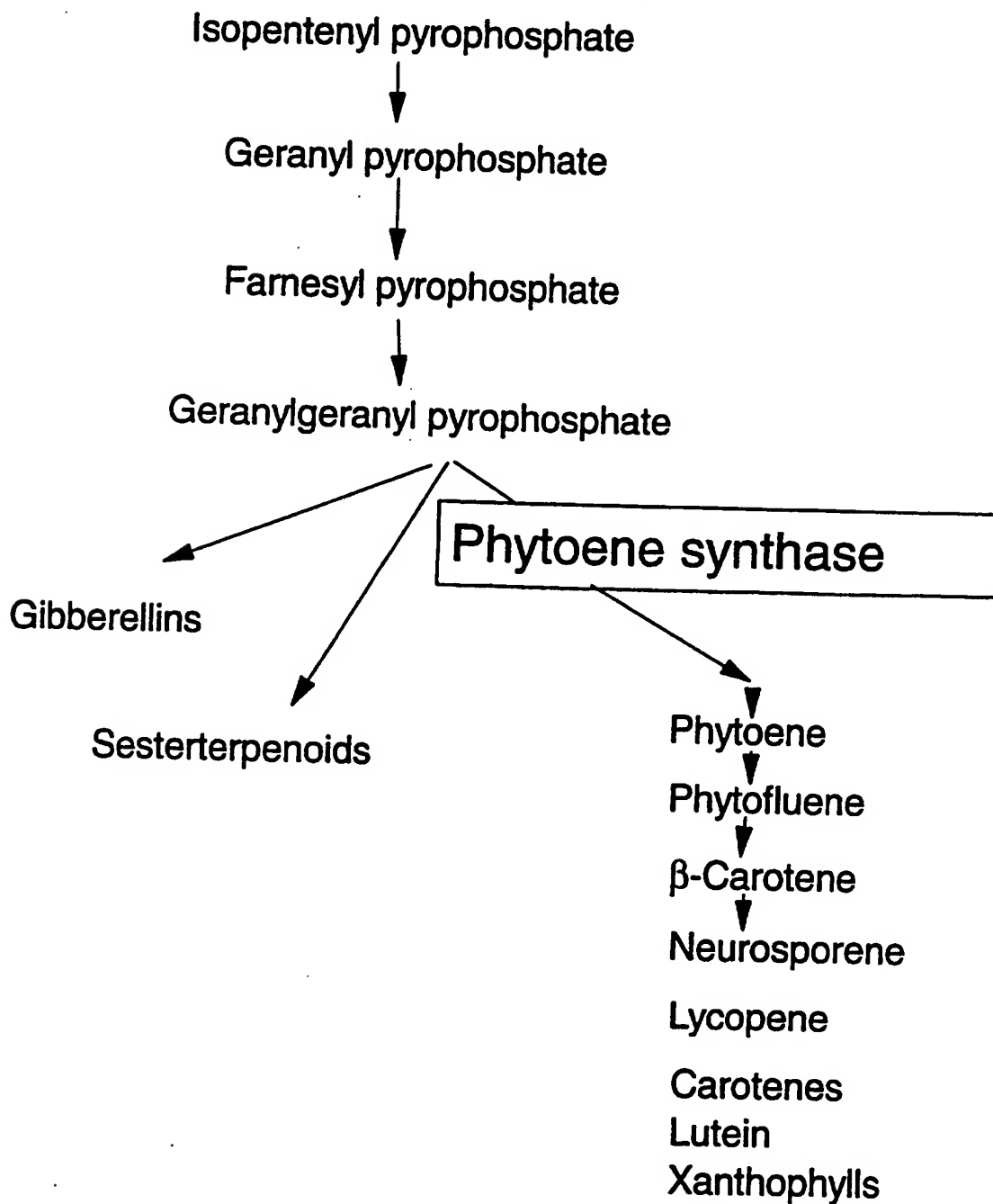
CLAIMS

1. A DNA encoding melon phytoene synthase.
2. The cDNA encoding melon phytoene synthase having the nucleotide sequence ID-1 .
3. A DNA construct comprising some or all of a DNA sequence encoding melon phytoene synthase under the control of a transcriptional initiation region operative in plants, so that the construct can generate RNA in plant cells.
4. A DNA construct as claimed in claim 3, in which the said DNA sequence is in antisense orientation.
5. A DNA construct as claimed in claim 4, in which the said DNA is a full length equivalent of the melon phytoene synthase gene.
6. A DNA construct as claimed in claim 4, in which the said DNA is a partial sequence containing a sufficient number of nucleotides to effect inhibition of the expression of the endogenous phytoene synthase gene of a plant.
7. A DNA construct as claimed in claim 3, in which the said DNA is in sense orientation.
8. A DNA construct as claimed in claim 7, in which the said DNA is a full length equivalent of the melon phytoene synthase gene.
9. A DNA construct as claimed in claim 7, in which the said DNA is a partial sequence of the melon phytoene synthase gene.

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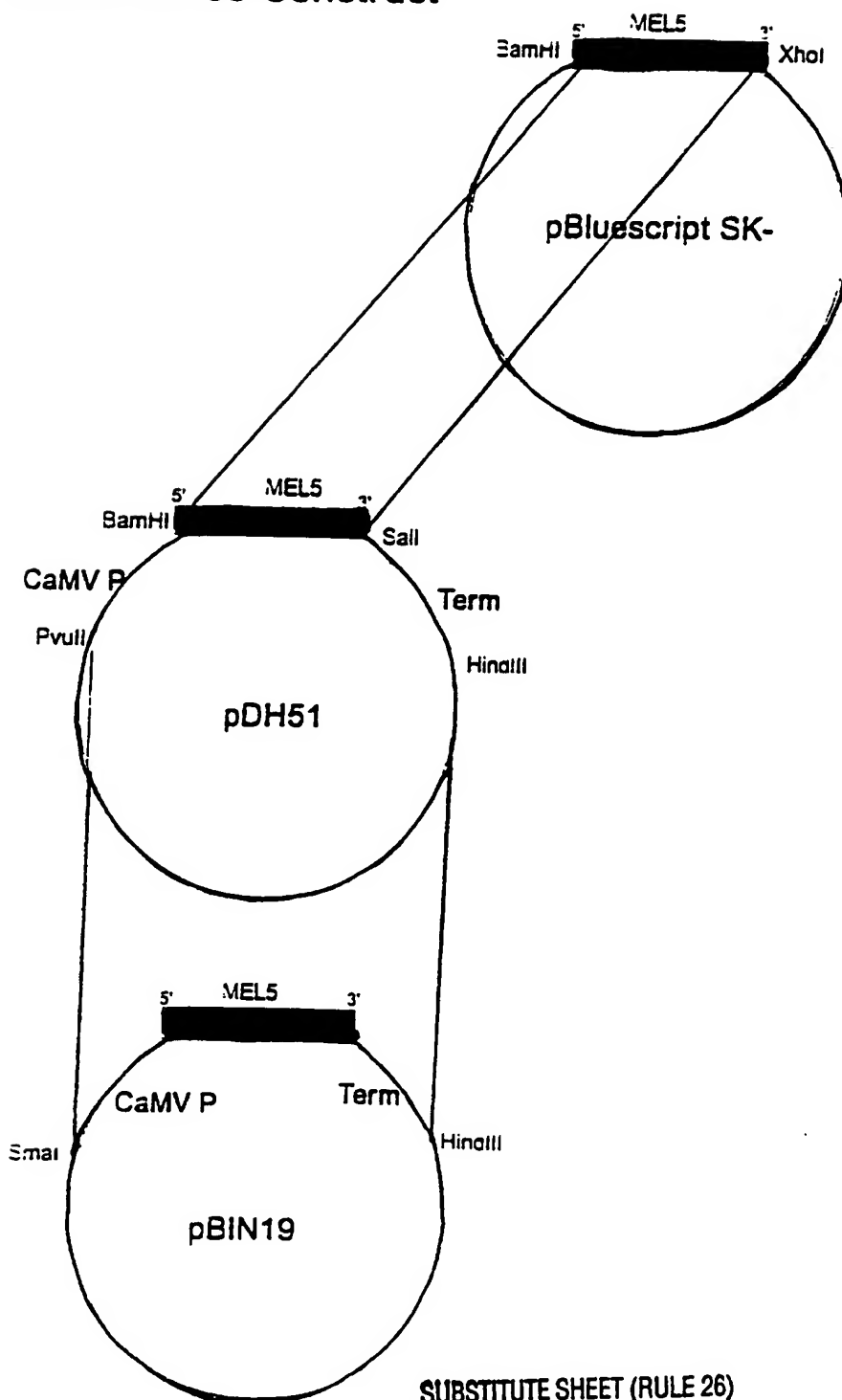
10. A DNA construct as claimed in any of claims 3 to 9, in which said DNA comprises a sequence of at least 10 nucleotides for transcription into RNA.
11. A DNA construct as claimed in claim 10, in which the said DNA comprises at least 35 nucleotides.
12. A method of modifying the production of carotenoids in a plant comprising transforming said plant with DNA adapted to modify carotenoid biosynthesis and growing said transformed plants or their descendants followed by selection of plants having plant parts of modified carotenoid content.
13. A method as claimed in claim 12, in which the plant is modified to enhance the production of one or more carotenoids or to inhibit such production by the plant.
14. A plant have a modified ability to produce carotenoids.

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Figure 1**CAROTENOID BIOSYNTHESIS**

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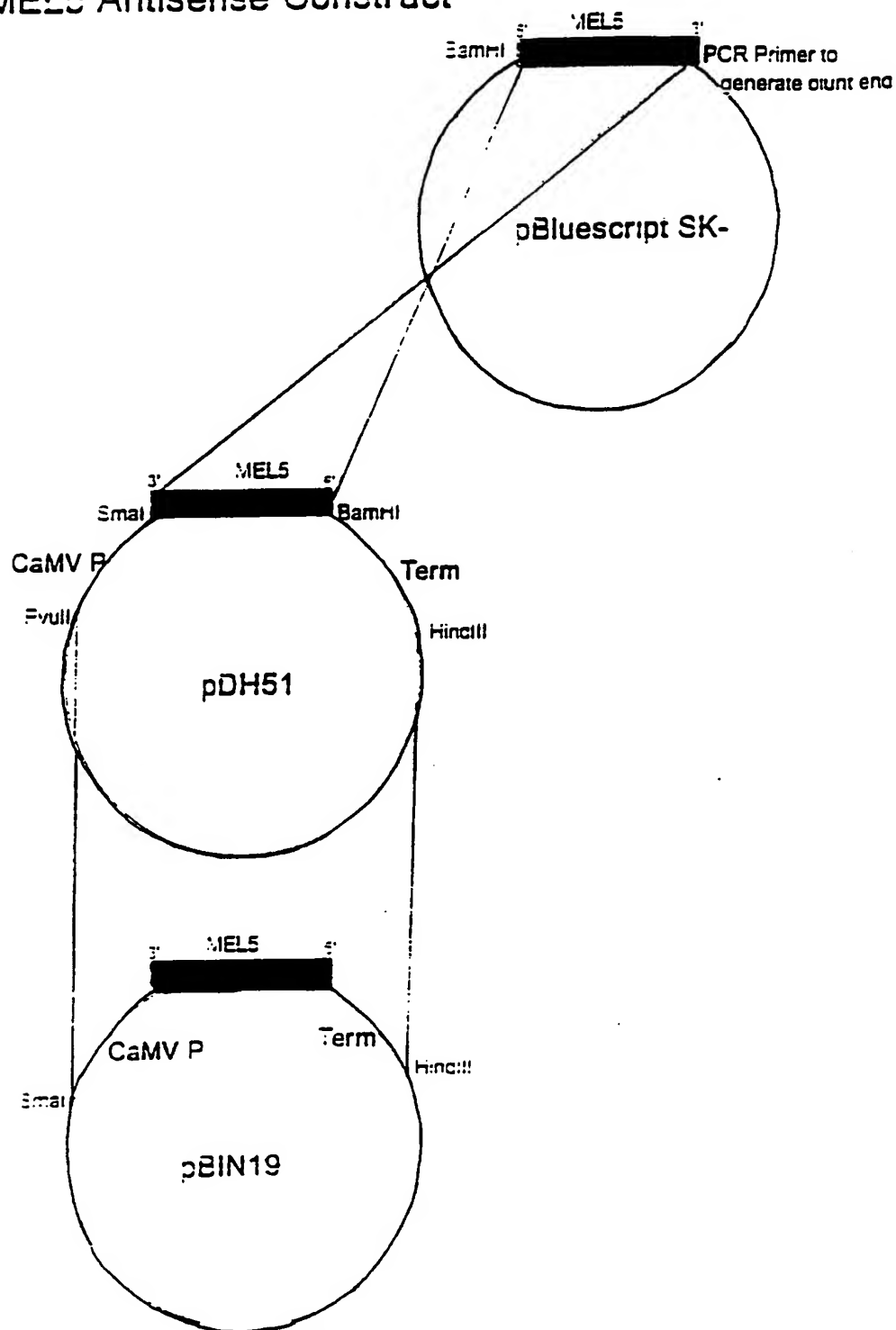
Figure 2
MEL5 Sense Construct



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Figure 3

MEL5 Antisense Construct



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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/GB95/01603 (22) International Filing Date: 6 July 1995 (06.07.95) (30) Priority Data: 9414505.9 18 July 1994 (18.07.94) GB 9419081.6 22 September 1994 (22.09.94) GB (71) Applicant (for all designated States except US): ZENECA LIMITED [GB/GB]; 15 Stanhope Gate, London W1Y 6LN (GB). (72) Inventors; and (75) Inventors/Applicants (for US only): KARVOUNI, Zoi [GR/GR]; Vioryl, 36 Viltaniotis Street, GR-145 64 Kifissia (GR). JOHN, Isaac [PK/GB]; University of Nottingham, Dept. of Physiology and Environmental Science, Sutton Bonington, Loughborough LE12 5RD (GB). TAYLOR, Jane [GB/GB]; Lancaster University, Institute of Environmental and Biological Sciences, Division of Biological Sciences, Lancaster LA1 4YQ (GB). WATSON, Colin [GB/US]; University of Arizona, Dept. of Plant Sciences, Tucson, AZ 85721 (US). TURNER, Andrew [GB/GB]; Scotgen Limited, 2 Lillydron Avenue, Aberdeen AB9 2TN (GB). GRIERSON, Donald [GB/GB]; University of Nottingham, School of Agriculture, Dept. of Plant	Physiology, Sutton Bonington, Loughborough LE12 5RD (GB). (74) Agents: HUSKISSON, Frank, Mackie et al.; Zeneca Agrochemicals, Intellectual Property Dept., Jealott's Hill Research Station, P.O. Box 3538, Bracknell, Berkshire RG12 6YA (GB). (81) Designated States: AM, AU, BB, BG, BR, BY, CA, CN, CZ, EE, FI, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LV, MD, MG, MN, MW, MX, NO, NZ, PL, RO, RU, SD, SG, SI, SK, TJ, TM, TT, UA, UG, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG). Published <i>With international search report.</i> (88) Date of publication of the international search report: 13 February 1997 (13.02.97)	
(54) Title: DNA, CONSTRUCTS, CELLS AND PLANTS DERIVED THEREFROM (57) Abstract A DNA encoding melon phytoene synthase.		

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INTERNATIONAL SEARCH REPORT

International Application No PC1/GB 95/01603		
A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/52 C12N15/82 A01H5/00		
According to International Patent Classification (IPC) or to both national classification and IPC		
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Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	THE PLANT JOURNAL, vol. 2, no. 3, 1993 pages 343-349, BRAMLEY, P., ET AL. 'BIOCHEMICAL CHARACTERIZATION OF TRANSGENIC TOMATO PLANTS IN WHICH CAROTENOID SYNTHESIS HAS BEEN INHIBITED THROUGH EXPRESSION OF ANTISENSE RNA TO PTOM5' see the whole document <div style="text-align: center;">--- -/--</div>	10-14
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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X	WO,A,91 09128 (ICI PLC) 27 June 1991 see the whole document ---	10-14
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Information on patent family members

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